



0271-5198(94)00054-9

HEMORHEOLOGIC EFFECTS OF LIGHT PROLONGED EXERCISE

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(Received 21.07.1994; accepted 09.08.1994)

ABSTRACT

Increases in blood viscosity during various kinds of strenuous exercise have been repeatedly described. In this study we investigated the rheologic effects of long duration low intensity exercise. Twelve volunteers (21-39 yr, 6 men and 6 women), after overnight fasting, performed a 60 min exercise on cycloergometer at 55% of the theoretical maximal heart rate. After an early increase at the 10th minute ($p < 0.001$), blood lactate decreases ($p < 0.02$) and returns to normal. During exercise there is an increase in plasma viscosity ($p < 0.001$) and hematocrit ($p < 0.05$) at the 10th minute. Red cell rigidity index "Tk" increases at the 20 th minute ($p < 0.05$). Whole blood viscosity ($p < 0.01$) increases and hematocrit/viscosity ratio decreases ($p < 0.01$). Thus, a light prolonged work load induces a transient hyperviscosity pattern very similar to that which is observed during strenuous exercise bouts.

Key words: Blood viscosity, hematocrit, exercise, hemorheology, erythrocyte deformability, blood lactate.

INTRODUCTION

Increased physical activity is now considered as an important tool against cardiovascular disease (1-3). While strenuous exercise is sometimes at risk for acute cardiovascular accidents (4), low intensity prolonged exercise is considered as a safe procedure for improving both lipidic and glucidic metabolism and reducing atherogenetic abnormalities (5). Such a 'metabolic fitness' may be clearly obtained by light muscular activity, such as walking at a brisk pace, which represents 50% of the maximal aerobic power (6). Many kinds of exercise have been shown to acutely impair blood rheology (7-12) but we were not aware of studies concerning light prolonged exercise and rheology. Therefore, we investigated the possible hemorheologic effects of 1 hr cycling at 55% of the maximal theoretical heart rate.

SUBJECTS AND METHODS

Exercise test

Twelve healthy volunteers (age: 21-39 yr, 6 men and 6 women) remained fasting until they performed at 9 a.m. the exercise test. An indwelling catheter was set in a superficial vein of the cubital fossa. Blood samples were drawn at -15, 0, 10, 20, 30, 40, 50, 60 min as well as 10 min after stopping exercise (time 70 min). Exercise-tests were performed on a bicycle ergometer (Bodyguard, Jonas Oglænd A.S., N 4301- Sandnes, Norway). Heart rate was continuously monitored with the impulses coming from three electrodes taped to the subject's chest. Subjects exercised 1 hr at 55% of the maximal theoretical heart rate given by the tables of the American Heart Association.

Hemorheological measurements

Blood samples for hemorheological measurements (7 ml) were obtained with a large bore needle (Luer adaptor Venoject, set into the catheter) to avoid shear damage to erythrocytes. A vacuum tube was used for blood withdrawal, with potassium EDTA as the anticoagulant. No tourniquet was used for sample drawing in order to minimize venous stasis. Viscometric measurements were performed at high shear rate (1000 s^{-1}) with a falling ball viscometer (MT 90 Medicatest, 37 rue de l'Ermitage F-86280 Saint Benoit) (13-15). Accuracy of the measurements was regularly controlled with the Carrimed Rheometer 'CS' (Rhéo, 19 rue Ambroise Croizat, 91120 Palaiseau, France). The coefficient of variation of this method ranges between 0.6 and 0.8% (10 repetitive measurements of the same sample). We measured with this device apparent viscosity of whole blood at native hematocrit, plasma viscosity, and blood viscosity at corrected hematocrit (45%) according to the equation of Quemada (16):

$$\mu_b = \mu_{pl} \cdot (1 - 1/2 k \cdot h)^{-2}$$

where μ_s is blood viscosity, μ_{pl} plasma viscosity, h the hematocrit and k a shear dependent intrinsic viscosity of the red cells according to Quemada.

Two indices of erythrocyte rigidity (Dintenfass' 'Tk' and Quemada's 'k') were calculated from blood viscosity, hematocrit and plasma viscosity measured at time 0 with equations derived from those given above:

$$k = 2 \cdot (1 - \mu_r - 0.5) \cdot h^{-1}$$

and:

$$Tk = (\mu_r^{0.4} - 1) \cdot (\mu_r^{0.4} \cdot h)^{-1} \quad (17)$$

Where μ_r is relative blood viscosity μ_b/μ_{pl} . The hematocrit/viscosity ratio, an index of oxygen supply to tissues, was calculated according to Chien (18) and Stoltz (19), with h (as percentage) divided by μ_b value at high shear rate which was determined as described above.

Biochemical analyses

The sampled blood was centrifuged and the plasma assayed for diverse parameters by well standardized and routine techniques, on an automatic clinical analyzer (DuPont de Nemours). Both lactate and ammonia were assayed with the kits from DuPont specially adapted to this analyzer. Blood lactate assay was based on NADH production by rabbit lactate dehydrogenase. Coefficients of variation range between 0,7 and 5.6 %.

Changes in plasma volume

A formula for calculating plasma volume changes (% ΔPV) during exercise from hematocrit changes has been published by investigators of the NASA-Ames Research Center (20,21,22) who demonstrated its validity in moderate as well as maximal exercise. We applied this formula to our data. The equation is:

$$\% \Delta PV = 100 / (100 - H_o) \times 100 [(H_o - H) / H_o]$$

where H_o is resting hematocrit and H hematocrit during exercise.

Statistics.

Results are presented as mean \pm the SE of the mean. Correlations were performed using the method of least squares. Variables were compared using the nonparametric test of Mann-Whitney and Wilcoxon. Significance was defined as $p < 0.05$. The choice of nonparametric tests was done in order to

adhere the guidelines of the ICSH expert panel for blood rheology (23), since hemorheological parameters usually appear to exhibit a nonnormal distribution.

RESULTS

For reasons discussed elsewhere (15), viscosity values obtained with the MT90 viscometer are 1.3 fold lower than those measured at the same shear rate with the Carri-Med Rheometer. In this study, as well as our other ones, we did not apply this correction factor. Comparison of our values with values that would be given by a rotational viscometer can be made by multiplying the results by 1.3.

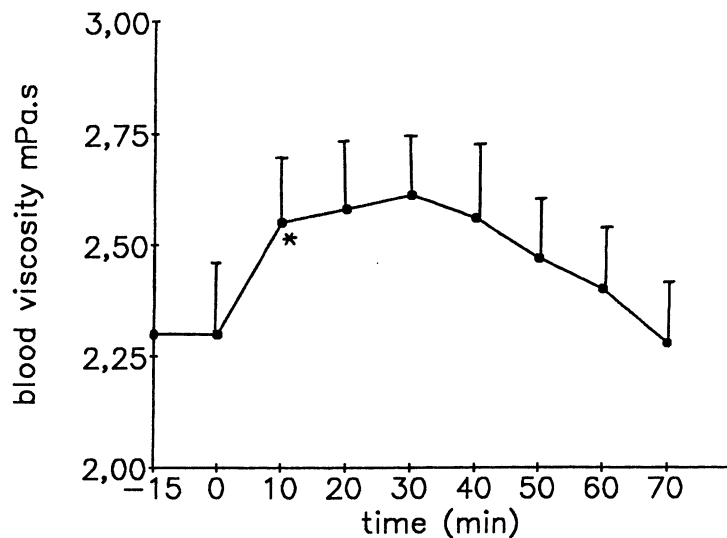


FIG.1

*Effects of light prolonged exercise on whole blood viscosity at native hematocrit: * $p < 0.05$ vs baseline.*

Fig. 1 shows that blood viscosity increases during the test ($p < 0.05$ at time 10 min). This increase is explained by an increase in hematocrit (fig. 2), plasma viscosity (fig.3), and RBC rigidity (fig. 4). Blood viscosity at corrected hematocrit 45% is also increased (+ 9.5% $p < 0.01$). The hematocrit/viscosity ratio is decreased at t 10 (-5.8 % $p < 0.01$). Blood lactate increases from baseline values of 1.83 ± 0.22 mmol/l up to 2.57 ± 0.29 mmol/l at 10 min and then gradually decreases (fig. 5) to return within the resting range after 40 min. Plasma volume changes calculated with the equation of Greenleaf showed a

reduction of this volume (-6 to -7%) during all the session, followed by a normalization at the 10th minute of recovery (fig. 6).

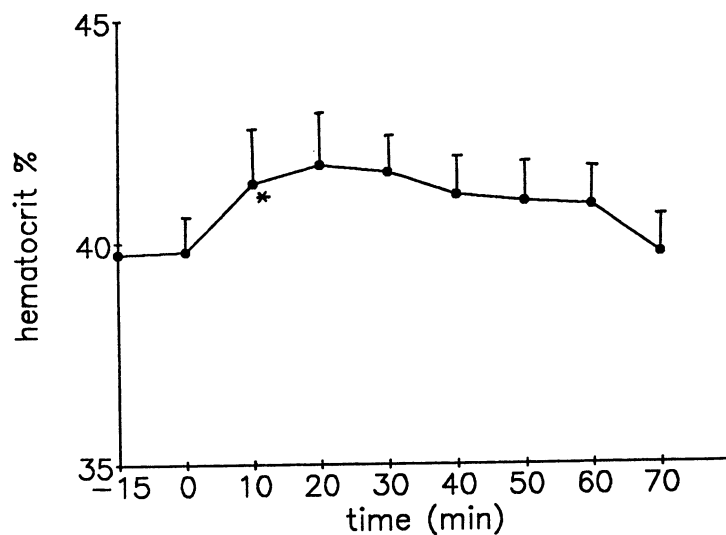


FIG.2

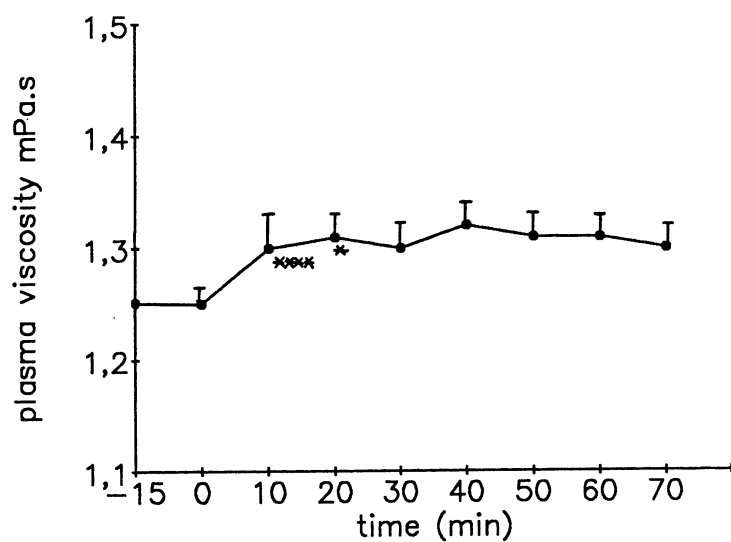
*Effects of light prolonged exercise on hematocrit. * $p < 0.05$ vs baseline.*

There is a correlation between Tk and blood lactate ($r=0.227$ $p<0.03$) as shown on fig. 7.

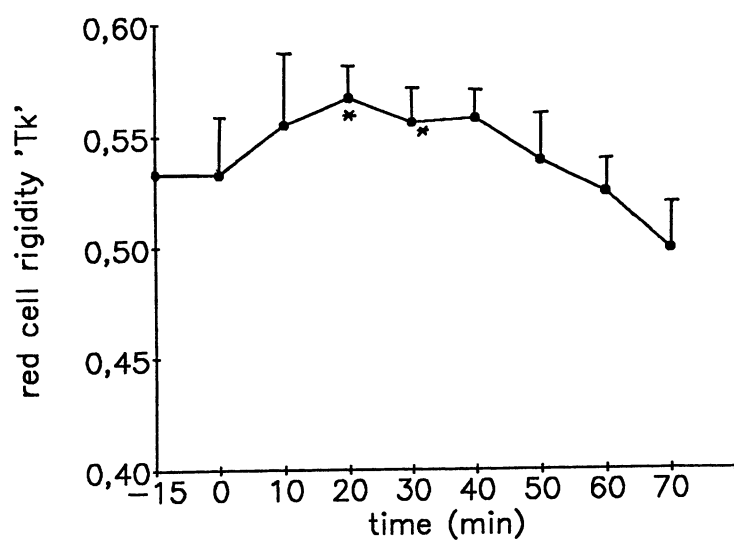
DISCUSSION

This study shows that very light exercise induces the same hemorheologic changes which have been reported during strenuous work loads. A rise in hematocrit, which indicates an outward movement of fluid from the vascular bed to the interstitial space during muscular activity (22) is a classical finding, even with a low intensity exercise (21). It was more surprising to notice that when viscosity was corrected for hematocrit, its increased remained highly significant, due to a rise in both plasma viscosity and red cell rigidity.

Since the red cell rigidity index "Tk" is correlated with blood lactate measured throughout the test, a role for blood lactate in this increase in red cell rigidity can be hypothesized, as previously published for stronger exercise protocols (12, 24, 25). Since both lactate (26) and acidosis (27) reduce red cell deformability, such a mechanism is theoretically possible. However, the increase in lactate observed during this protocol was moderate, far below the onset of metabolic

**FIG.3**

Effects of light prolonged exercise on plasma viscosity . * $p < 0.05$
 *** $p < 0.01$ vs baseline.

**FIG.4**

Effects of light prolonged exercise on RBC rigidity index 'Tk' . * $p < 0.05$ vs baseline.

acidosis (28). This kind of exercise is considered as an 'aerobic' one in which fatigue is more related to glycogenic depletion and hypoglycemia, while lactate synthesis from pyruvate is not strongly stimulated (29). An early peak in blood lactate above 2 mmol/l is probably due to a rapid glucose oxidation to pyruvate, while the Krebs cycle becomes more slowly efficient (29). When the metabolism reaches a steady state, lactate is no longer released, as indicated by its progressive decline.

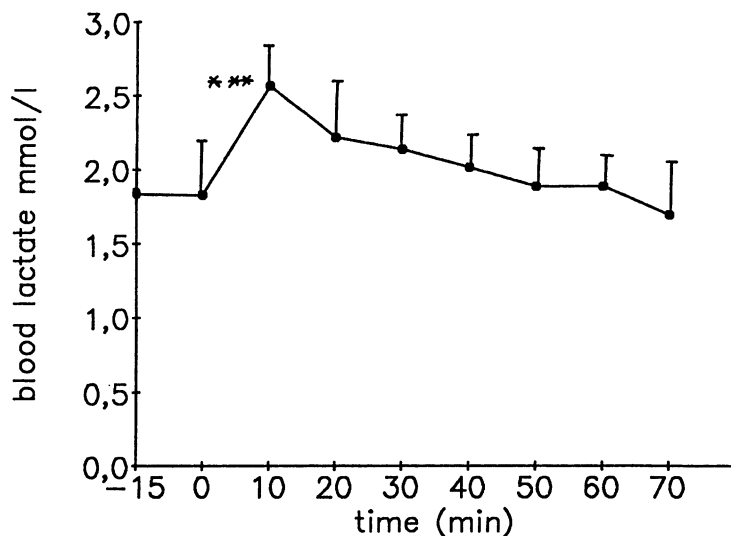


FIG.5

*Effects of light prolonged exercise on blood lactate. *** $p < 0.01$ vs baseline.*

Another mechanism which may improve blood rheology during prolonged exercise is the oxidative stress induced by increased free radicals production (30). However, the kinetics of our modifications of 'Tk' which appear early and progressively return towards normal preexercise values, does not support this hypothesis since oxidative stress is more likely to progressively increase with the duration of exercise. Clearly, 'Tk' changes appear to be more parallel to the transient lactate peak and we believe that the two phenomena are related.

An interesting hypothesis has been proposed by M. Gueguen-Delamaire (9) who suggested that impairment of blood rheology may be involved in the cardiovascular risk of maximal exercise, together with changes in hemocoagulatory parameters. Since we observe during this light, very safe exercise quite the same rheologic changes than during strong work loads, we think that these simple changes in hematocrit, red cell rigidity, and plasma viscosity are physiological adaptative modifications which occur during many

hematocrit/viscosity ratio, which measures the contribution of blood rheology to O₂ supply (18-19) is slightly decreased, but such a change can be easily overcome by vasodilatation. In our opinion, the risk of strong maximal or exhausting work loads is probably more related to modifications of hemostasis and to white cell activation.

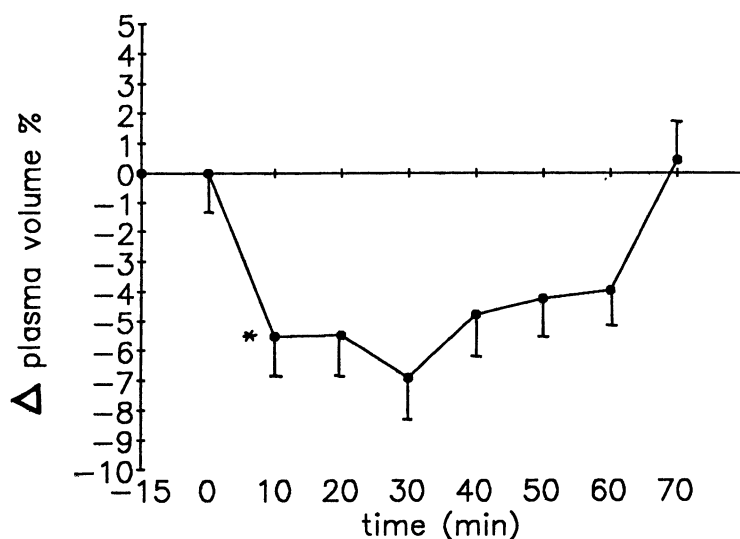


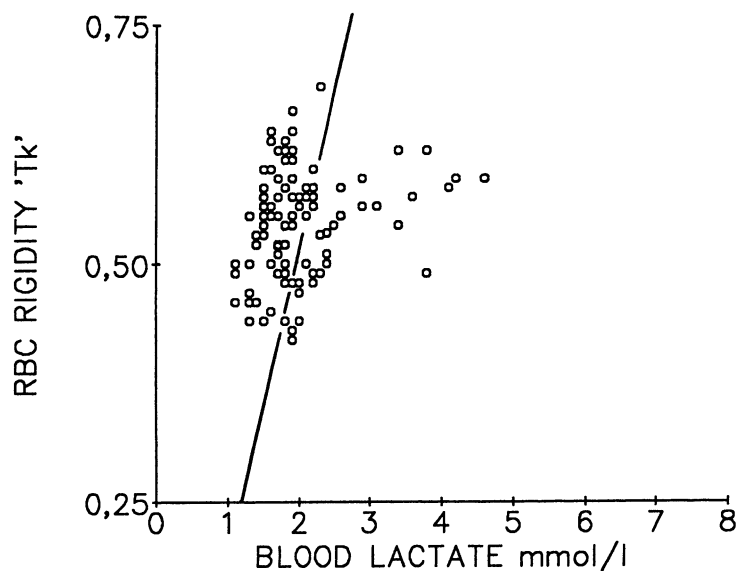
FIG.6

Changes in plasma volume calculated with the equations of Greenleaf during the prolonged light exercise session.

In conclusion, light prolonged exercise protocols at 55 % of the theoretical maximal heart rate induce the same hemorheologic modifications as strong short work loads, i.e. an increase in blood viscosity explained by a rise in hematocrit, plasma viscosity and erythrocyte rigidity. The latter event, which is transient and rapidly returns to normal, is correlated with blood lactate and may be related to its modifications during exercise.

ACKNOWLEDGEMENT

Our thanks are extended to Negma pharmaceuticals for their kind financial and technical support.

**FIG.7**

Correlation between Tk and blood lactate during the prolonged light exercise session. $r=0.227$ $p<0.03$.

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